

Cell Certificate of Analysis - Amended

Product Description	WA26	WA26	
Cell Line Provider	WiCell	WiCell	
Parent Material	This material descended from derivation	This material descended from derivation	
Lot Number	WB0131	WB0131	
Date Vialed	26-February-2012		
Passage Number	p8 ¹	p8 ¹	
Culture Platform	Feeder Independent	Feeder Independent	
	Medium: E8 plus PVA	Matrix: Recombinant Human Vitronectin	

The following testing specifications have been met for the specified product lot.

Test Description	Test Provider	Test Method	Test Specification	Result
Post-Thaw Viable Cell Recovery	WiCell	SOP-CH-305	≥ 15 Undifferentiated Colonies, ≤ 30% Differentiation	Pass ²
Identity by STR	UW Molecular Diagnostics Laboratory	PowerPlex 16 HS System by Promega	Consistent with known profile	Pass
Sterility - Direct transfer method	Apptec	30744	Negative	Pass
Mycoplasma	Charles River	ID 91/0	Negative	Pass
Karyotype by G-banding	WiCell	SOP-CH-003	Normal karyotype	Pass

¹These cells were cultured for 7 passages prior to freeze. WiCell adds +1 to the passage number at freeze so that the number on the vial best represents the overall passage number of the cells at thaw. Footnote provided by TL on 17Sep12.

The following tests were performed on the cell line. The tests do not apply to any particular lot. Please see the individual test reports for results of each test.

Test Description	Test Provider	Test Method
Differentiation Potential by Teratoma	WiCell	SOP-CH-213 SOP-CH-214
HLA	UW Histocompatibility Laboratory	High resolution sequencing method with Celera reagents on the ABI 3100 instrument
ABO	New York Blood Center	For ABO: Olsson ML, Chester MA. A rapid and simple ABO genotype screening method using a novel B/O2 versus A/O2 discriminating nucleotide substitution at the ABO locus. Vox Sang 1995; 69(3):242-7. For RHD: Singleton BK, Green CA, Avent ND, Martin PG, Smart E, Daka A, Narter-Olaga EG, Hawthorne LM, Daniels G. The presence of an RHD pseudogene containing a 37 base pair duplication and a nonsense mutation in Africans with the Rh D-negative blood group phenotype. Blood 2000; 95(1): 12-8.
Growth Curve (Doubling Time)	WiCell	Varies by culture platform
Flow Cytometry for ESC Marker Expression	WiCell	SOP-CH-024
Comprehensive Human Virus Panel	Charles River	ID 91/0

²Post-Thaw Viable Cell Recovery was obtained using Rho-Kinase Inhibitor (Y-27632) during thaw.



Certificate of Analysis - Amended

Cells distributed by WiCell are intended for research purposes only and are not intended for use in humans.

Appropriate biosafety precautions should be followed when working with these cells. The end user is responsible for ensuring that the cells are handled and stored in an appropriate manner. WiCell is not responsible for damages or injuries that may result from the use of these cells.

Amendment(s):

Reason for Amendment	
CoA updated to include copyright information.	See Signature
Amended STR test method and HLA test provider and test method.	
Original CoA.	

Date of Lot Release	Quality Assurance Approval
16-August-2012	AMC AMC Quality Assurance Signed by:



Short Tandem Repeat Analysis*

Sample Report: 10502-STR

Label on Tube: 10502-STR

Sample Date: 05/11/12

Lab Received 05/11/12

Requestor: WiCell Research Institute

Test Date: 05/16/12

File Name: 120517

Report Date: 05/21/12

Sample Name: (label on tube) 10502-STR

Description: WI Cell Research Institute provided

genomic DNA

26.4 ug/mL 260/280=1.92

Locus	Repeat #	STR Genotype
D16S539	5, 8-15	12,14
D7S820	6-14	10,10
D13S317	7-15	11,12
D5S818	7-15	12,12
CSF1PO	6-15	11,11
TPOX	6-13	8,11
Amelogenin	NA	X,X
TH01	5-11	7,10
vWA	11, 13-21	17,17

Comments: Based on the 10502-STR DNA submitted by WI Cell dated and received on 05/11/12, this sample (Label on Tube: 10502-STR) exactly matches the STR profile of the human stem cell line WA26 comprising 12 allelic polymorphisms across the 8 STR loci analyzed. No STR polymorphisms other than those corresponding to the human WA26 stem cell line were detected and the concentration of DNA required to achieve an acceptable STR genotype (signal/ noise) was equivalent to that required for the standard procedure (~1 ng/amplification reaction) from human genomic DNA. This result suggest that the 10502-STR DNA sample submitted corresponds to the WA26 stem cell line and was not contaminated with any other human stem cells or a significant amount of mouse feeder layer cells. Sensitivity limits for detection of STR polymorphisms unique to either this or other human stem cell lines is ~5%.

5/22/12 Data

Date

Molecular Diagnostics Laboratory

55/a/

Molecular Diagnostics Laboratory

* Testing to assess engraftment following bone marrow transplantation was accomplished by analysis of human genetic polymorphisms at STR loci. This methodology has not yet been approved by the FDA and is for investigational use only.

File: Final STR Report

This report is confidential. No part may be used for advertising or public announcement without written permission. Results apply only to the sample(s) tested.



WiCell Research Institute

Report Number 900594 Page 1 of 1

June 12, 2012 P.O. #:

STERILITY TEST REPORT

Sample Information:

Stem Cells

1: WA27-WB0130 10504 7: 2: WA26-WB0131 10505 8:

3: WA25-WB0132 10506 4: WA25-WB0127 10507

5: WA26-WB0128 10508 6: WA27-WB0138 10509 7: WA26-WB0152 10514

8: WA25-WB0151 10512 9: WA09-WB0143 10521 10: WA09-WB0139 10520

11: WA27-WB0150 10522

12: H9 hOct4-pGZ-WB0140 10518 13: MIRJT6i-mND1-4-WB0142.10519

Date Received: Date in Test: Date Completed: May 23, 2012 May 29, 2012 June 12, 2012

Test Information:

Test Codes: 30744, 30744A Immersion, USP / 21 CFR 610.12 Procedure #: BS210WCR.201

TEST PARAMETERS	PROE	DUCT
Approximate Volume Tested	0.5 mL	0.5 mL
Number Tested	26	26
Type of Media	SCD	FTM
Media Volume	400 mL	400 mL
Incubation Period	14 Days	14 Days
Incubation Temperature	20 °C to 25 °C	30 °C to 35 °C
RESULTS	26 NEGATIVE	26 NEGATIVE

QA Reviewer

Date

Technical Reviewer

Date

Testing conducted in accordance with current Good Manufacturing Practices.



Charles River Research Animal Diagnostic Services

Sponsor: WiCell Research Institute Accession #: 2012-015912 Diagnostic Summary Report 20 Mar 2012 Received: Approved: 27 Mar 2012, 13:11 Bill Method: **Test Specimen:** Human ? **PDG** Sample Set Service (# Tested) **Profile Tested** +/-Assay #1 Infectious Disease PCR (3) All Results Negative + = Positive, +/- = Equivocal, ? = Indeterminate, PDG = Pending

	Service Approvals	
Service	Approved By*	Date
Infectious Disease PCR		27 Mar 2012, 13:11

To assure the SPF status of your research animal colonies, it is essential that you understand the sources, pathobiology, diagnosis and control of pathogens and other adventitious infectious agents that may cause research interference. We have summarized this important information in infectious agent **Technical Sheets**, which you can view by visiting http://www.criver.com/info/disease_sheets.

CR RADS ILIMS Form: FM-1741 Rev. 3

^{*}This report has been electronically signed by laboratory personnel. The name of the individual who approved these results appears in the header of this service report. All services are performed in accordance with and subject to General Terms and Conditions of Sale found in the Charles River Laboratories-Research Models and Services catalogue and on the back of invoices.

Product: Not Indicated

Charles River Research Animal Diagnostic Services

Sponsor: WiCell Research Institute

Test Specimen: Human Received: 20 Mar 2012

Accession #: 2012-015912

Molecular Diagnostics Infectious Disease PCR Results Report

Department Review: Approved by 27 Mar 2012, 13:11*

Human Comprehensive Virus Panel

Sample #: Code :	<u>1</u> WA25-WB0132 10429	<u>2</u> WA26-WB0131 10430	<u>3</u> WA27-WB0130 10431
John Cunningham virus	-	-	-
BK virus	-	-	-
Herpesvirus type 6	-	-	-
Herpesvirus type 7	-	-	-
Herpesvirus type 8	-	-	-
Parvovirus B19	-	-	-
Epstein-Barr Virus	-	-	-
Hepatitis A virus	-	-	-
Hepatitis B virus	-	-	-
Hepatitis C virus	-	-	-
HPV-16	-	-	-
HPV-18	-	-	-
Human T-lymphotropic virus	-	-	-
Human cytomegalovirus	-	-	-
HIV-1	-	-	-
HIV-2	-	-	-
Adeno-associated virus		-	-
Human Foamy Virus	-	-	-
LCMV PCR	-	-	-
Hantavirus Hantaan PCR	-	-	-
Hantavirus Seoul PCR	-	-	-
Mycoplasma Genus PCR	-	-	-
DNA Spike	PASS	PASS	PASS
RNA Spike	PASS	PASS	PASS
NRC	PASS	PASS	PASS

Remarks: - = Negative; I = Inhibition, +/- = Equivocal; + = Positive.

Sample Suitability/Detection of PCR Inhibition:

Sample DNA or RNA is spiked with a low-copy number of a exogenous DNA or RNA template respectively. A spike template-specific PCR assay is used to test for the spike template for the purpose of determining the presence of PCR inhibitors. The RNA spike control is also used to evaluate the reverse-transcription of RNA. Amplification of spike template indicates that there is no detectable inhibition and the assay is valid.

NRC:

The nucleic acid recovery control (NRC) is used to evaluate the recovery of DNA/RNA from the nucleic acid isolation process. The test article is spiked with a low-copy number of DNA/RNA template prior to nucleic acid isolation. A template-specific PCR assay is used to detect the DNA/RNA spike.

^{*}This report has been electronically signed by laboratory personnel. The name of the individual who approved these results appears in the header of this service report.



Chromosome Analysis Report: 008056

Report Date: May 17, 2012

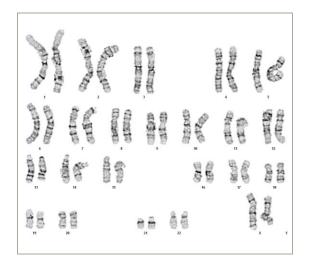
Cell Line: WA26-WB0131 10502

Passage #: 10

Date of Sample: 5/4/2012

Date Completed: 5/17/2012

Results: 46,XX



Specimen: hESC on rh Vitronectin

Cell Line Gender: Female

Reason for Testing: pre-freeze

Investigator:

, WiCell

Derivation

Cell: S01-22

Slide: 1(10)KARYOTYPE
Slide Type: Karyotyping

of Cells Counted: 20

of Cells Karyotyped: 4

of Cells Analyzed: 8

Band Level: 450-625

Interpretation:

No clonal abnormalities were detected at the stated band level of resolution.

Completed by	, CG(ASCP), on 5/17/2012	
Reviewed and interpreted by		PhD, FACMG, on 5/17/2012

A signed copy of this report is available upon request.

-3	
Date:	Sent To:
Sent By:	QC Review By:

Limitations: This assay allows for microscopic visualization of numerical and structural chromosome abnormalities. The size of structural abnormality that can be detected is >3-10Mb, dependent upon the G-band resolution obtained from this specimen. For the purposes of this report, band level is defined as the number of G-bands per haploid genome. It is documented here as "band level", i.e., the range of bands determined from the four karyograms in this assay. Detection of heterogeneity of clonal cell populations in this specimen (i.e.,mosaicism) is limited by the number of metaphase cells examined, documented here as "# of cells counted".

This assay was conducted solely for listed investigator/institution. The results may not be relied upon by any other party without the prior written consent of the Director of the WiCell Cytogenetics Laboratory. The results of this assay are for research use only. If the results of this assay are to be used for any other purpose, contact the Director of the WiCell Cytogenetics Laboratory.



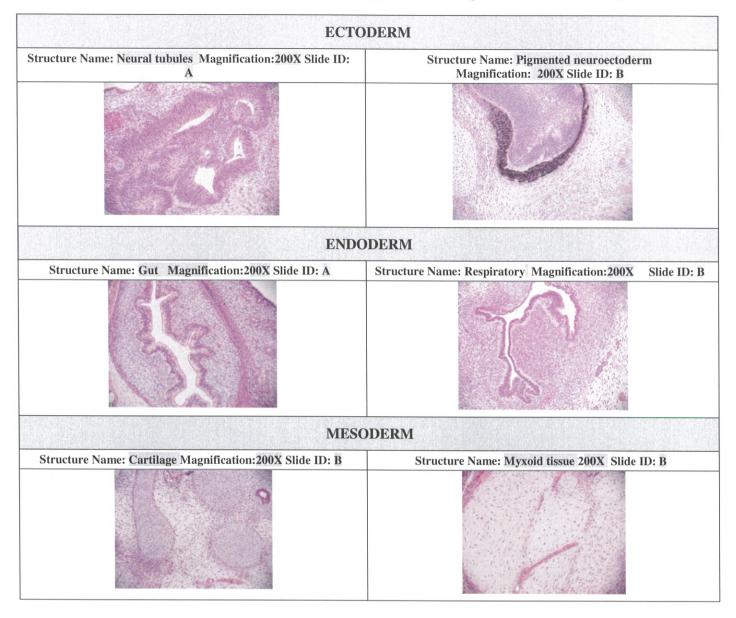


FORM SOP-CH-014.01 Version B Edition 01

Cell Line: WA26

Cell Lot Number: NA

Sample Number: 10405-A,B,C



Comments: Structures identified include Ectoderm (2), Mesoderm (2) and Endoderm (2)

Sample(s) were assessed for the presence of differentiation into cell types characteristic of the three embryonic germ layers, which, if present in the sample(s) examined, are represented in the photographs above. The individual's signature below verifies that this report accurately reflects the pathology observed.

Pathologist (By/Date): 6/27/2012

QA Review (By/Date): __

Print Date: 28-Jun-12



University of Wisconsin Hospital and Clinics

Name:

WICELL, 10405 HLA

MRN:

OS000183

Hospital: Physician: ,

Category:

DOB: HLA#:

WICELL

Bone Marrow Case Histocompatibility Summary 301417-DT

HLA Typing Results

Patient

Relation Hap A*

<u>C*</u>

DRB1*

DRB3* DRB4* DRB5* DQB1* DPB1*

The following allele combination(s), in which both alleles are listed by the ASHI CWD review committee as

Tested Date Collect Date

03/12/12

WICELL, 10405 HLA OS000183 / WICELL Patient

02:01 02:01

51:01

07:02:01G 07:01

18:01:01G 07:01:01G 01:01

03/01/12

HLA typings performed by sequencing, SSO, SSP or a combination. For low-resolution testing, results are reported by Serologic Equivalents. A "+" in the HLA allele designation indicates that the typing was performed by low/mid-resolution molecular method and that additional alleles are possible. Only the most frequent allele is listed.

HLA DNA-Based Typing

<u>Name</u>									
HLA / MR#	Method								
Received	Test Date	<u>A*</u>	<u>B*</u>	<u>C*</u>	DRB1*	DRB3*	DRB4*	DRB5*	DQB1*
WICELL, 10405_HLA		02:01							
OS000183 / WICELL	SEQ	02:01							
03/01/2012	03/20/2012								
		HLA Alle	le database:	IMGT 3.7.0 2	2012-01-12				
			18:01:0	1G					
	SEQ		51:01						
03/01/2012	03/20/2012								
1		antigen r	ecognition s	roup B*18:01: ite of exons 2 IMGT 3.7.0 2	and 3: B*18	ne following all 3:01 B*18:17		nare identical	sequences in the
				07:01:0)1G				30
	SEQ			07:02:0)1G				
03/01/2012	03/20/2012								
		HLA Allele database: IMGT 3.7.0 2012-01-12 The reported allele group C*07:01:01G includes the following alleles, which share identical sequences in the							
		The repo	orted allele g		01G includes th		eles, which sl	hare identical	sequences in the
		antigen r	ecognition s	ite of exons 2	and 3: C*0	7:02 C*07:50	():		
					01:01				
	SEQ				07:01				
03/01/2012	03/20/2012								

HLA Allele database: IMGT 3.7.0 2012-01-12

Comments

Printed Date: 03/25/2012 UWHC 301417-DT

rare or not well defined, cannot be excluded: DRB1*01:21,07:11



University of Wisconsin Hospital and Clinics

Name:

WICELL, 10405 HLA

MRN:

OS000183

DOB:

HLA#: WICELL

Hospital:

Physician:

Category:

Bone Marrow Case Histocompatibility Summary

301417-DT

This test was developed and its performance characteristics determined by this laboratory. It has not been cleared or approved by the U.S. Food and Drug Administration. The FDA has determined that such clearance or approval is not necessary. This test is used for clinical purposes. It should not be regarded as investigational or for research. This laboratory is certified under the Clinical Laboratory Improvement Amendments of 1988 (CLIA-88) as qualified to perform high complexity clinical laboratory testing.

Electronically signed by

Director or Delegate, HLA Laboratory

03/25/2012 12:15

Date/Time

-43

Histocompatibility Laboratory, Room D4/231, 600 Highland Ave., Madison, WI 53792-2472 Teresa Darcy, MD, Medical Director :: Thomas M. Ellis, PhD, D(ABHI) Laboratory Director Lab: 608.263.8815 (option 3); Fax: 608.263.9610 ASHI: 01-4-WI-03-2, CLIA: 52DO661997

Printed Date: 03/25/2012 UWHC 301417-DT

▲ New York Blood Center

Laboratory of Immunohematology and Genomics 45-01 Vernon Blvd., Long Island City, N.Y. 11101 718-752-4771 • Fax 718-752-4747

March 20, 2012

WiCell Research Institute

SAMPLE: DNA WA26 #10405 (MA#167-12)

Date Received: 03/08/12 Sample Date: 03/01/12

HISTORY: DNA from cell line.

TESTING REQUESTED: Genotype for ABO and common RH

TESTING PERFORMED: *ABO*: Polymerase chain reaction-restriction fragment length polymorphism (PCR-RFLP) testing for nucleotide (nt) positions 261 (O¹), 467 (A²), 703 (B), and 1096 (B and O²). *RH*: Multiplex PCR-RFLP for *RHD* and *RHCE*C/c*. PCR-RFLP for RHCE Exon 5 (676C>G for E/e).

DNA MOLECULAR RESULTS: ABO: PCR-RFLP testing indicates the sample is homozygous for deletion of G at 261 characteristic of O^1 alleles. RH: RHD exons 4 and 7 are present. Negative for the inactivating RHD pseudogene. RH*Cc and RH*ee.

Genotype

Predicted Phenotype

WA26 #10405:

 $ABO*O^{1}O^{1}$; RH*D, RH*Cc, RH*ee

Group O; RhD+, C+E-c+e+

Manager, Genomics

Director of Immunohematology and Genomics

These *in vitro* diagnostic tests were developed and their performance characteristics established in the Molecular Analysis Laboratory. The tests have not been submitted to the Food and Drug Administration (FDA) for clearance or approval and; therefore, are not FDA-licensed tests. The Molecular Analysis Laboratory is certified under the Clinical Laboratory Improvement Amendment (CLIA) of 1988 as qualified to perform high complexity clinical testing. The New York Blood Center has been approved by the New York State Department of Health to perform these tests under its current Clinical Laboratory Permit.

These results are intended to predict a blood group antigen profile in a patient or donor, and are not intended for clinical diagnosis or as the sole means for patient management decisions. There are situations where testing DNA of a person may not reflect the red cell phenotype and not all performance characteristics have been determined. Nucleotide changes that inactivate gene expression or rare new variant alleles may not be identified in these assays. In addition, test results obtained from DNA isolated from leucocytes and other hematopoietic cells may differ from DNA isolated from other tissues in persons with a history of transplantation.

△ New York Blood Center

Immunohematology

Telephone: 718-752-4771

Genomics

Telephone: 718-752-4637

Sample: MA167-12; WA26 #10405

Test:

ABO and RH - GF

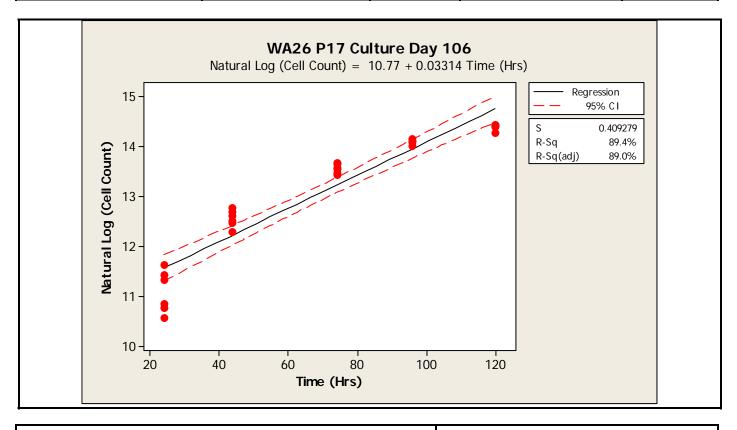
CPT CODE	Description/Molecular Testing	ABO/RH		
83892	Enzymatic digestion	X2		
83894	Separation by electrophoresis	x3		
83898	Amplification each nucleic acid seq	x3		
83912	Interpretation and report	X1		

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		4		



Characterization Report- Growth Characteristics

Sample ID	Cell Line	Cell lot #	Passage	Culture Day		Medium	Matrix	Passag	ing Additive
10449	WA26	N/A	17	7 106 E8 + PVA rh		rh-Vitronectin	Rho-kinase Inhibitor Y-27632		
Documentation of Growth Assay Data				Notebook #		Page(s)	Date Initiated		
					147		66-75	19APR12	
Growth Ass	Assay Performed by Report Prepared by Date QA Reviewed by		Date						
WiCell Derivation Laboratory LAN			14AUG12	JKT		15Aug12			



Regression Analysis: Natural Log (Cell Count) versus Time (Hrs)

The regression equation is Natural Log (Cell Count) = 10.8 + 0.0331 Time (Hrs)

 Predictor
 Coef
 SE Coef
 T
 P

 Constant
 10.7714
 0.1716
 62.75
 0.000

 Time (Hrs)
 0.033143
 0.002160
 15.35
 0.000

S = 0.409279 R-Sq = 89.4% R-Sq(adj) = 89.0%

Analysis of Variance Source DF SS MS F Ρ 0.000 Regression 39.452 39.452 235.52 **Residual Error** 28 4.690 0.168 44.143 Total 29

<u>Unusual Observations</u>

Time Natural Log Obs (Hrs) (Cell Count) Fit SE Fit Residual St Resid -0.9996 1 24 10.5672 11.5668 0.1270 -2.57R 5 10.7769 0.1270 -0.7899 -2.03R 24 11.5668 *R denotes an observation with a large standardized residual*.

Slope ± 95% C.I

 0.0331 ± 0.0044

Apparent Doubling Time (hours) ± 95% C.I.

20.91 ± 2.05

Apparent Doubling Time (95% C.I.)

18.45 hours - 24.14 hours



Procedure performed: Cell line: WA26

Passage 9

Sample ID: 10413

Date of: (03/05/12)

acquisition: file creation:

file submission:

	PERCENTS									
	SSEA4 -	SSEA4 +	SSEA4+	SSEA4 -	ALL	ALL				
antigen2:	antigen2 +	antigen2 +	antigen2 -	antigen2 -	SSEA4 +	antigen2 +				
SSEA3	0.01	44.10	55.50	0.32	99.60	44.11				
TRA1-60	1.05	91.00	7.69	0.23	98.69	92.05				
TRA1-81	2.96	87.50	8.69	0.85	96.19	90.46				
Oct-4	2.18	92.60	4.66	0.52	97.26	94.78				
SSEA1	0.02	3.98	95.70	0.33	99.68	4.00				

Percent analyzable events: 12.7

#wells submitted: 6

Total # cells analyzed: 2.26 X 10⁶

